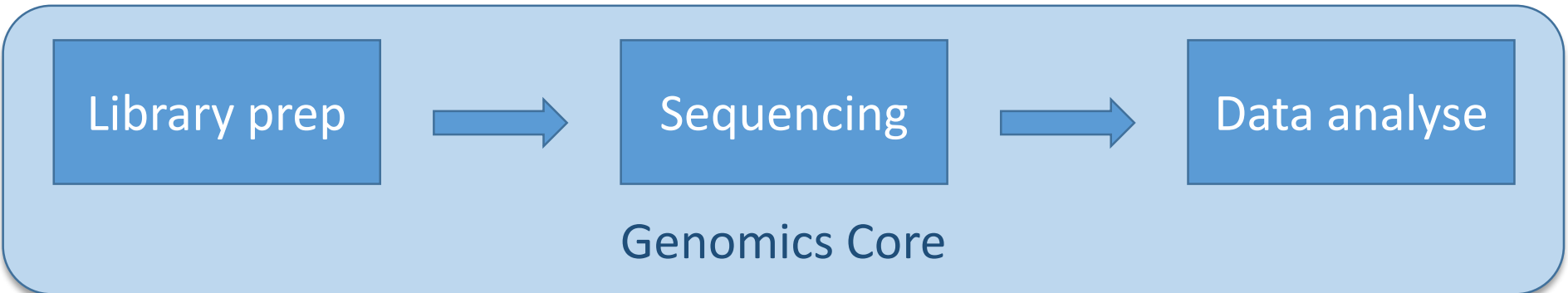
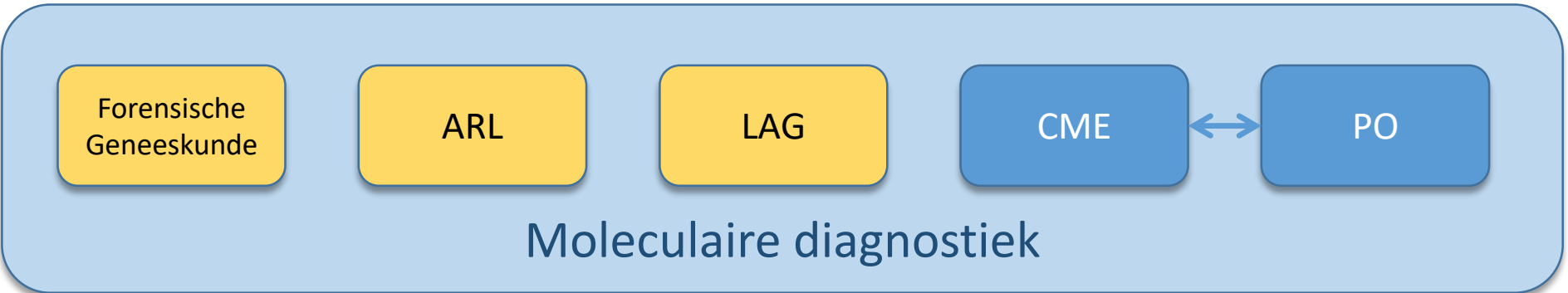


# NGS is microbiology

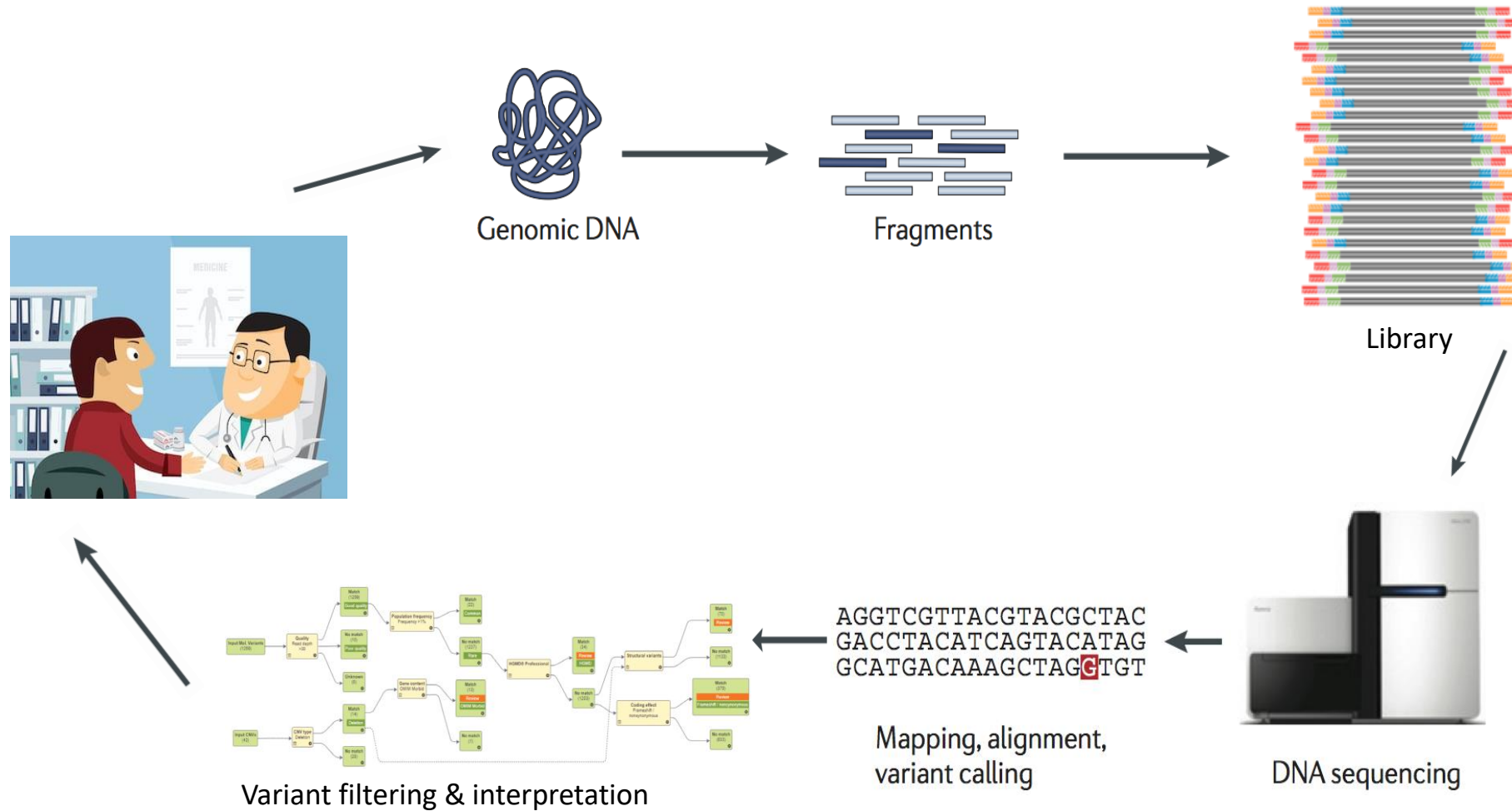
Wouter Bossuyt

Genomics Core, Leuven

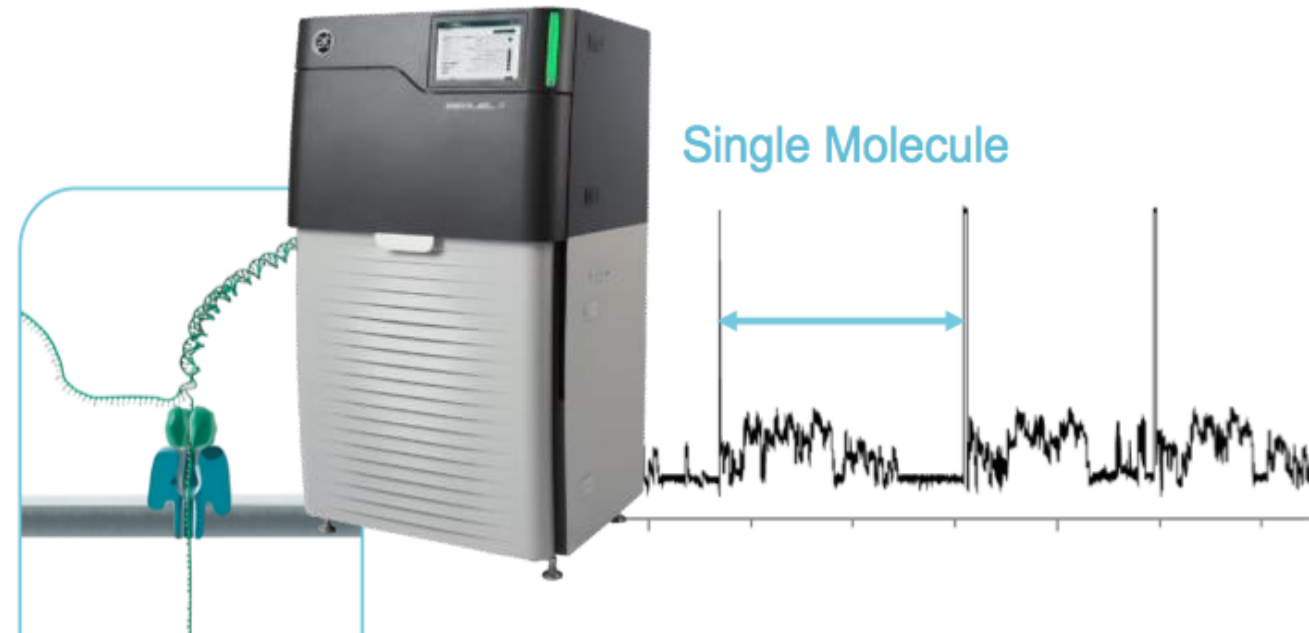
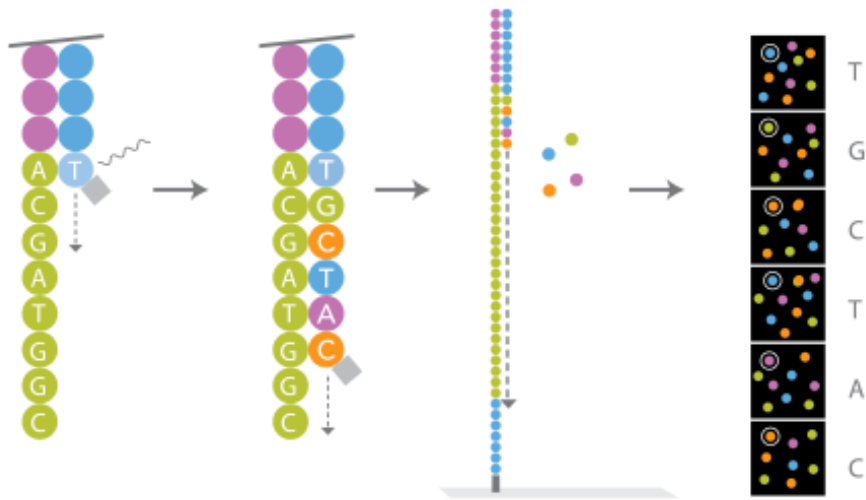
What's the use of technology if it is not usable?



# NGS workflow



# Two main technologies

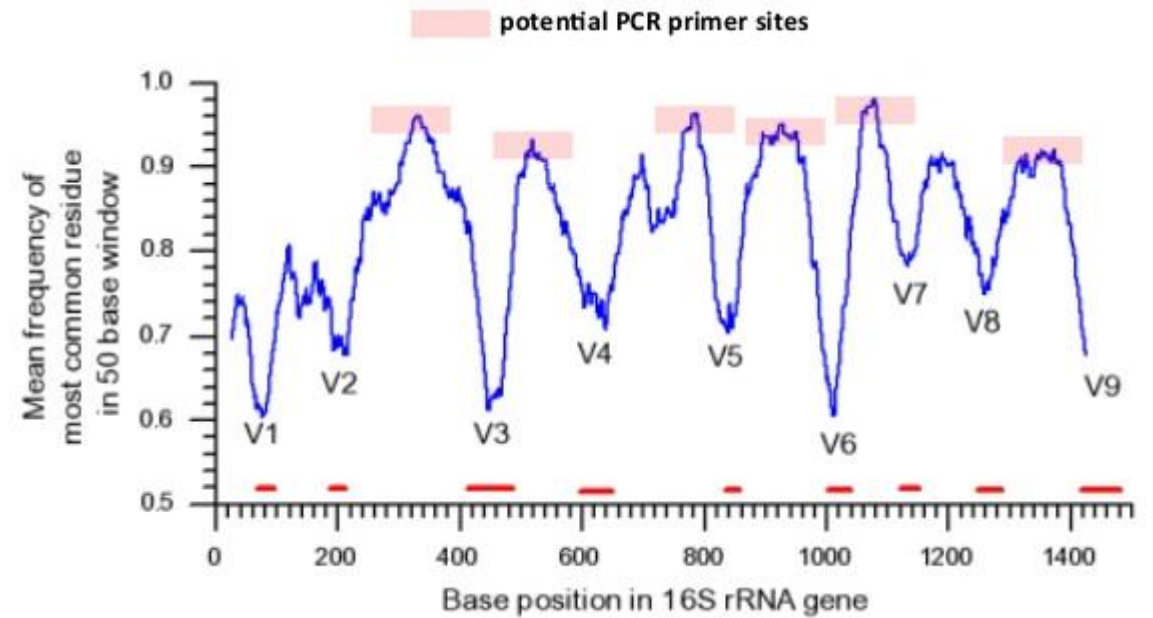
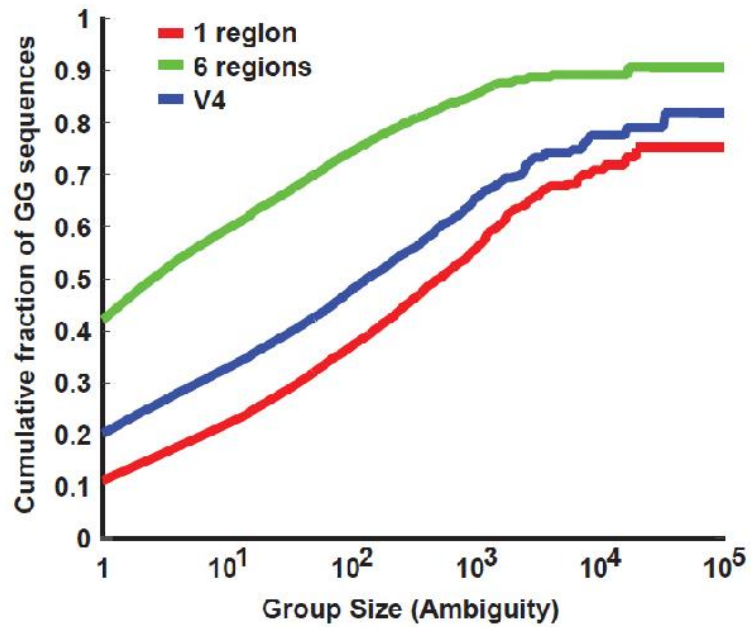


# applications

- Targeted amplicon sequencing
  - 16s
  - NGS MLST
- Bacterial WGS
- Metagenomics
- outbreak monitoring
  
- Virus discovery and detection

# Targeted approach: 16S RNA typing

- 16S typing
  - Fast analysis of microbial species in sample



# Targeted approach: NGS MLST

- Multilocus serotyping using NGS
- Multiplex PCR for 7 housekeeping genes → multiplex samples → sequence
  - Gene list dependent on taxonomy of interest
  - Sequence on long read sequencer
  - Depth of coverage important
- Limited info on resistance/virulence

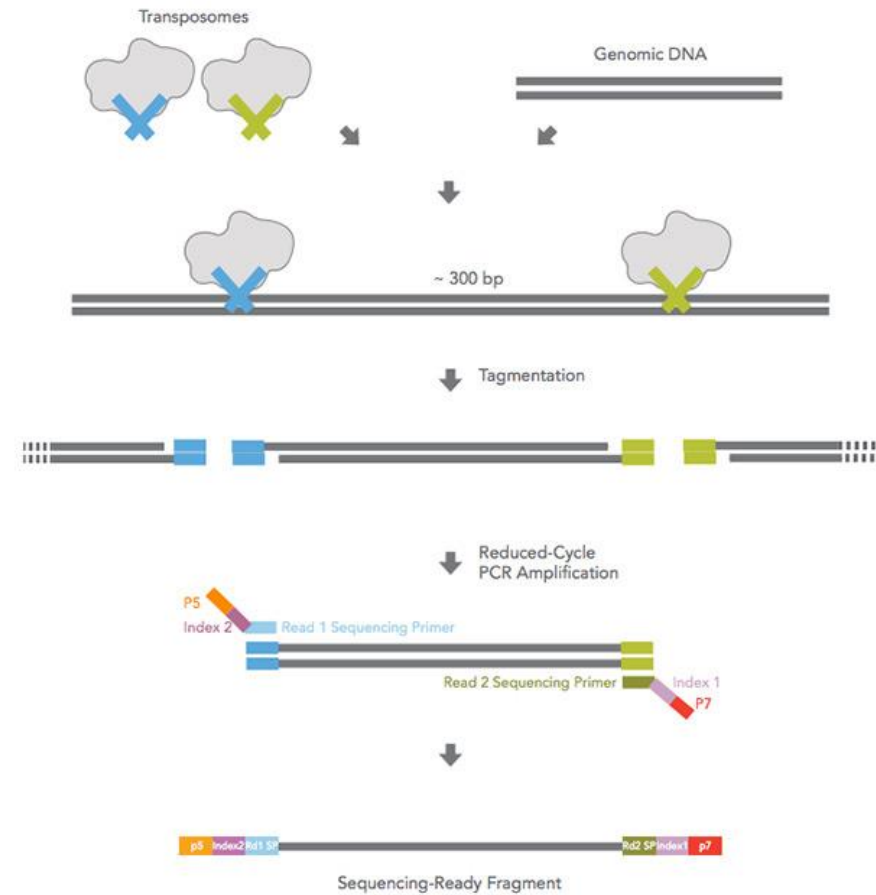
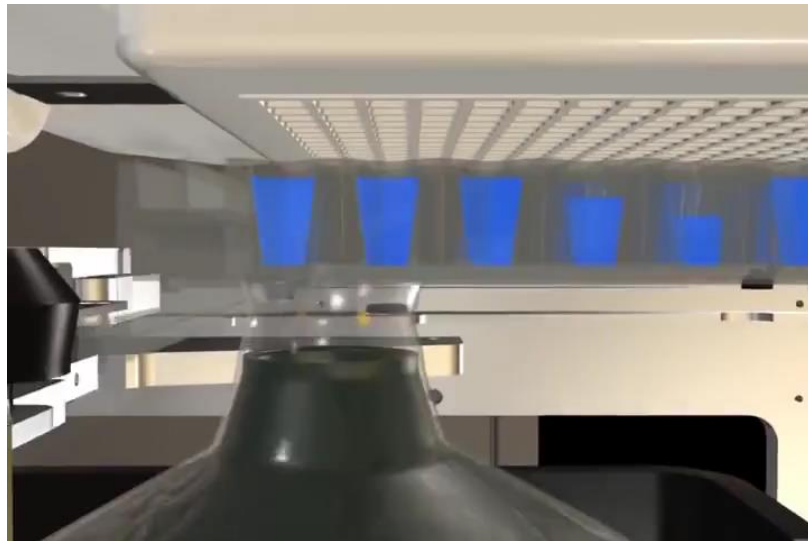


# Bacterial WGS

- Full sequencing of bacterial genomes
- Culture bacteria and isolate DNA
- Complete information
  - Typing
  - Pathogenecity monitoring during outbreak
  - Resistance monitoring
    - Identify known resistance mutations
    - Culture-based resistance monitoring in parallel to build more knowledge
  - *de novo* assembly to identify structural variants
  - Evolutionary tracking

# Automated library prep for bacterial WGS

- Nextera XT on Echo Labcyte
  - Proven method
  - Scaled to very low volumes
  - In 384-well plates

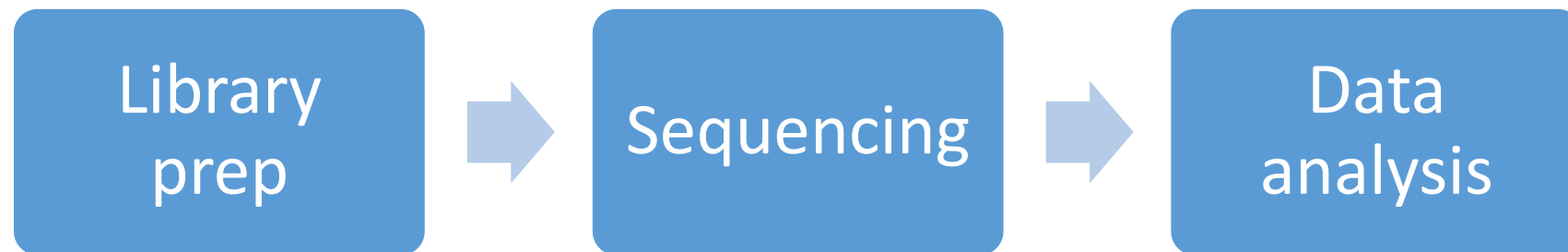


# Limited adoption of bacterial WGS

- Whole genome sequencing of bacterial genomes:
  - gives a lot of information
  - can combine many other tests in one test
- No large scale adoption
- What is holding people back?
  - Cost
  - Scalability
  - TAT
  - Knowhow

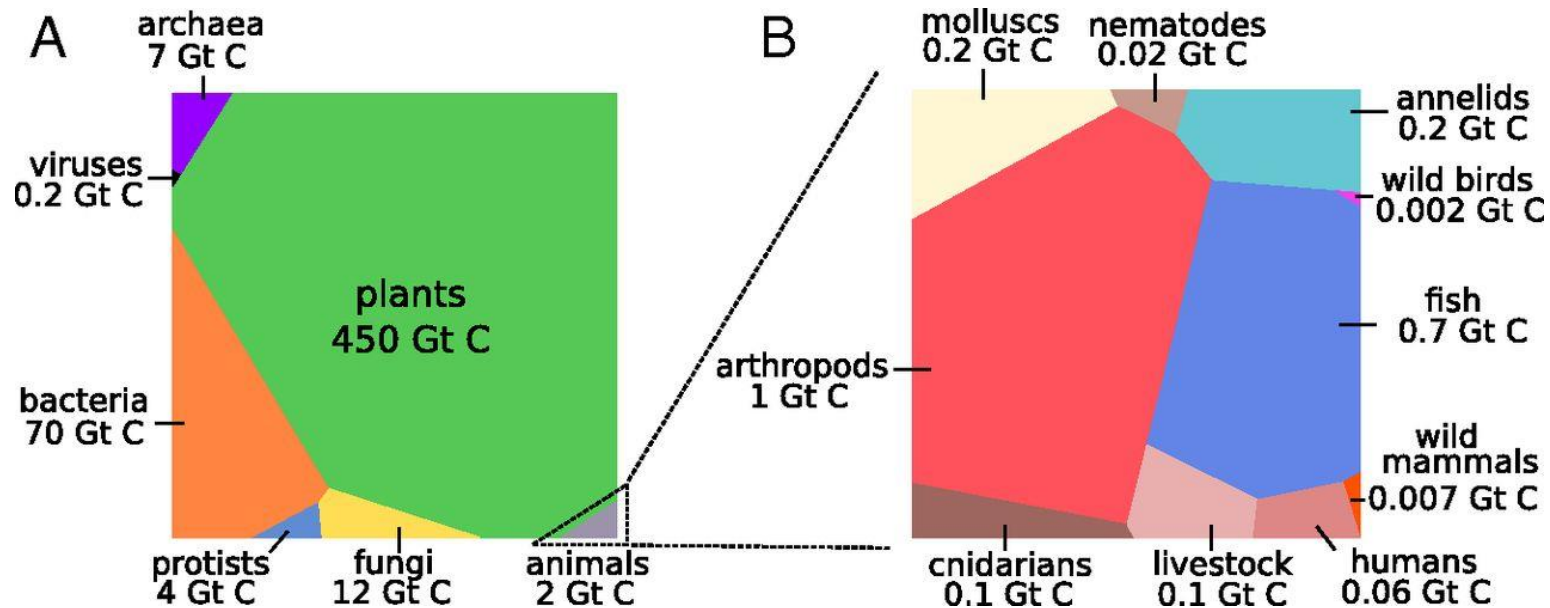
# Optimization of bacterial WGS workflow

- Possibility of automated sample submission
- LIMS integration
- Reservation and scheduling of necessary equipment
- Workflow optimization



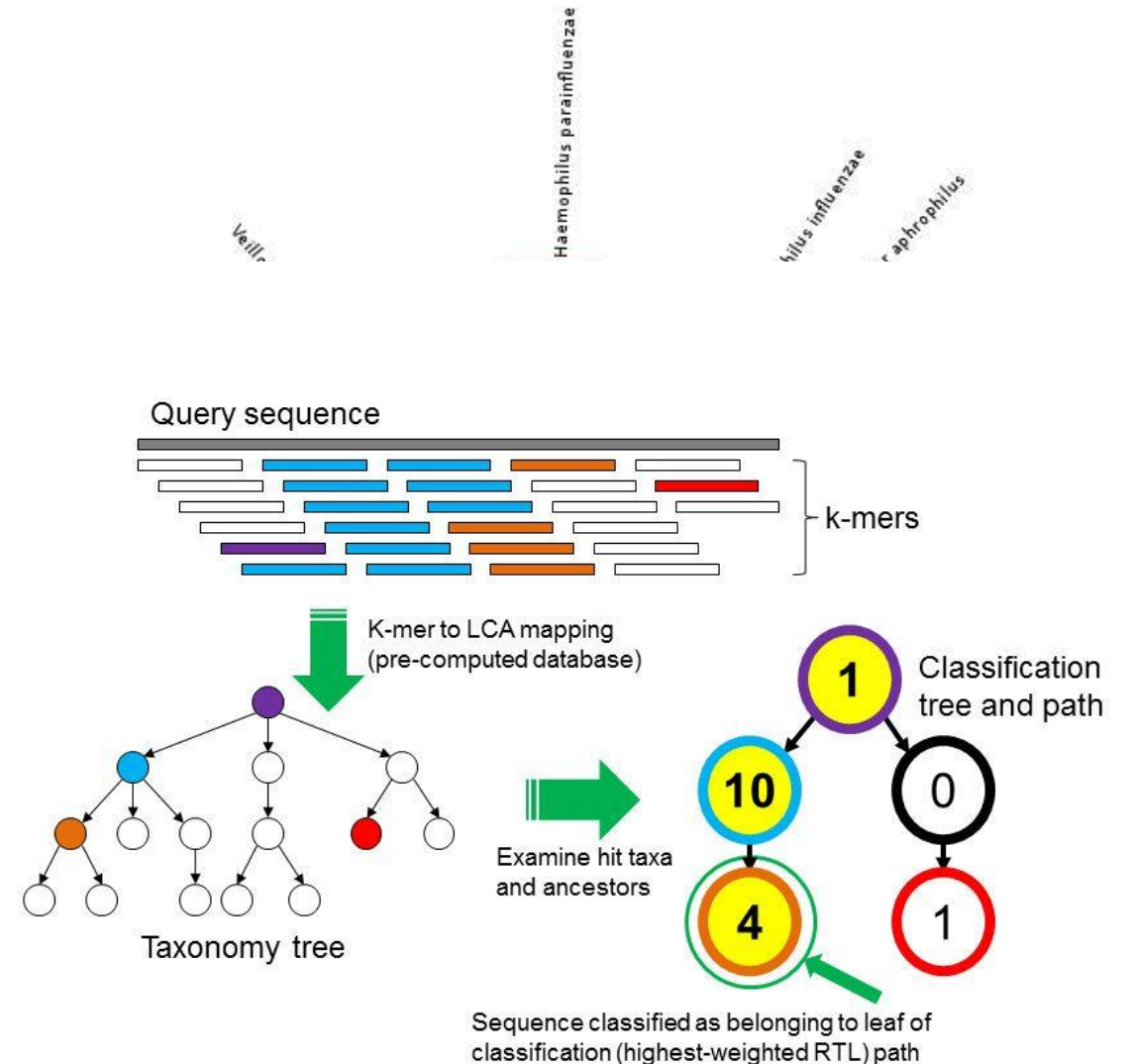
# Metagenomics

- Get DNA from a ROI → sequence
- Identification of different species in an 'environmental' sample



# Metagenomics

- Kraken as a tool for species identification
  - Identification of species
  - Quantification of species
- Possibility to select only reads from species of interest for further analysis

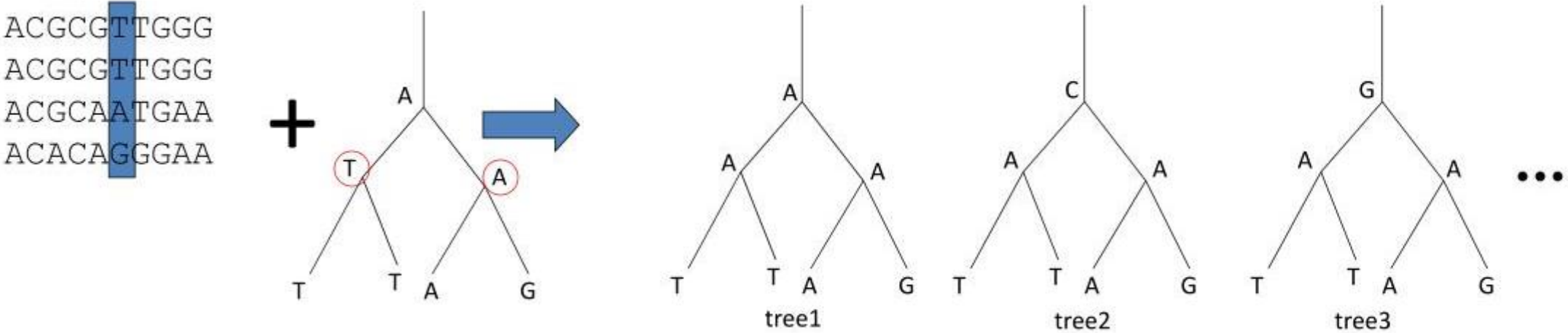


# Outbreak monitoring

## Metagenomics approach

1. Identify different bacterial species
  - Continue with likely species one at the time
2. Identify SNPs in sequence of closely related species
3. Use maximum likelihood approach to assemble phylogenetic tree

# Maximum likelihood



1. Probability is sum of probability of each branch
2. Calculate probability for each possible tree
3. Choose phylogenetic tree with maximum likelihood

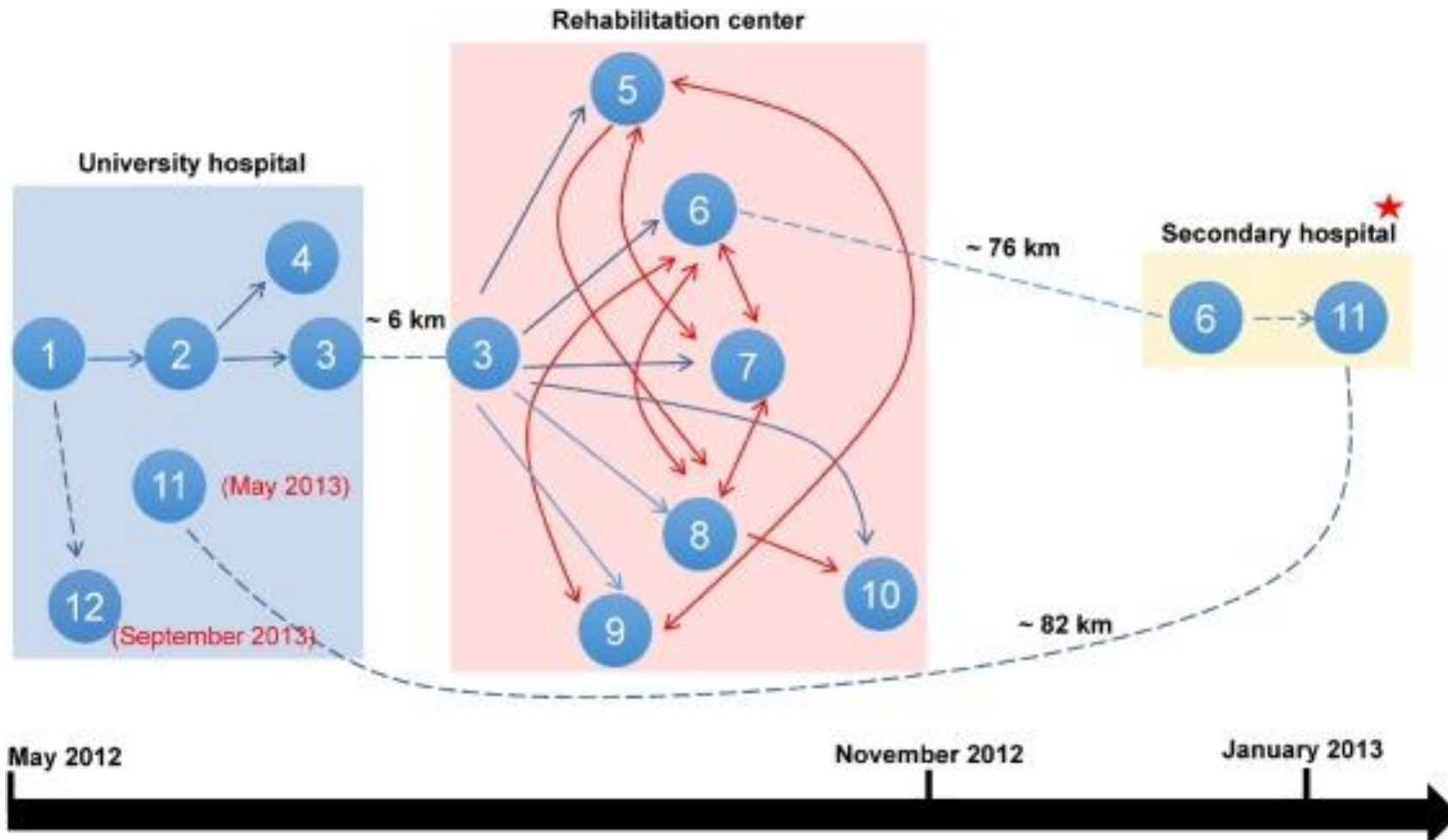


# Outbreak monitoring

## Metagenomics approach

1. Identify different bacterial species
  - Continue with likely species one at the time
2. Identify SNPs in sequence of closely related species
3. Use maximum likelihood approach to assemble phylogenetic tree
4. Align phylogenetic relationships with epidemiological evidence

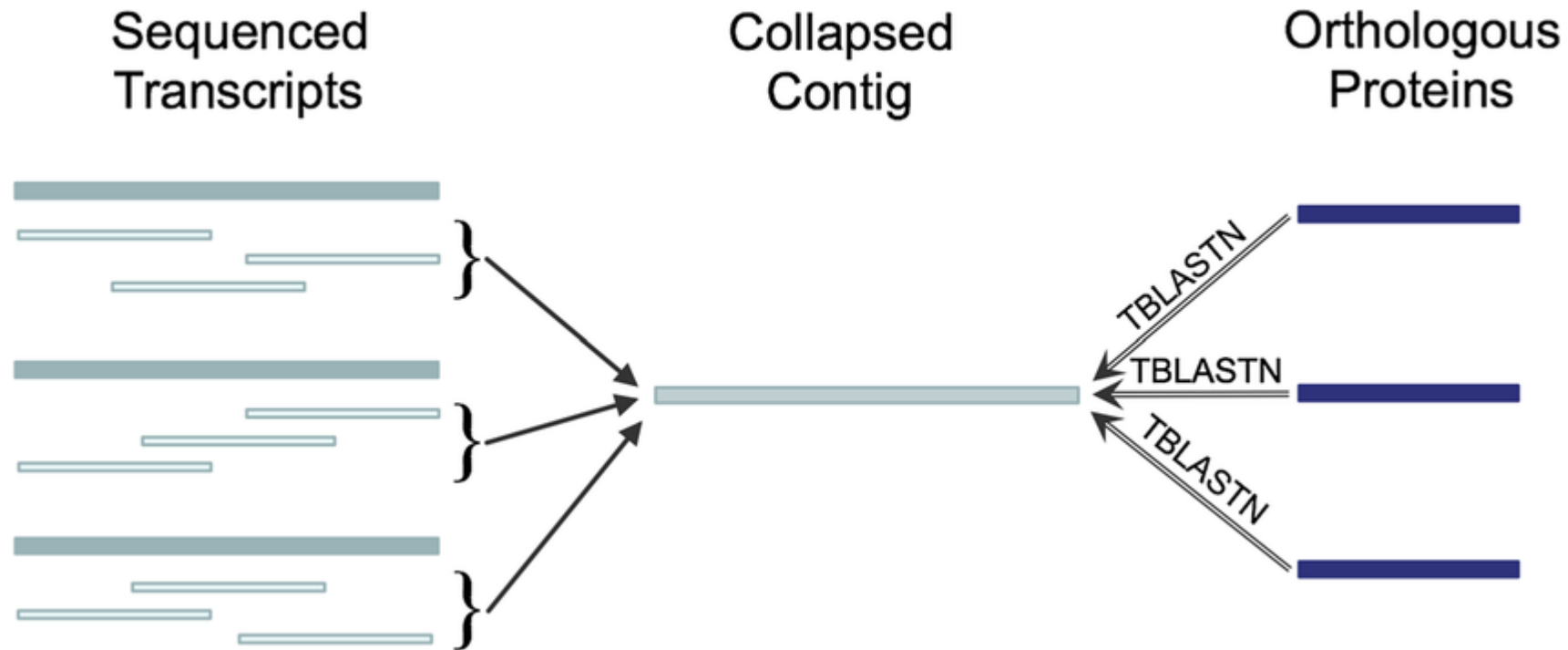
# Route of transmission



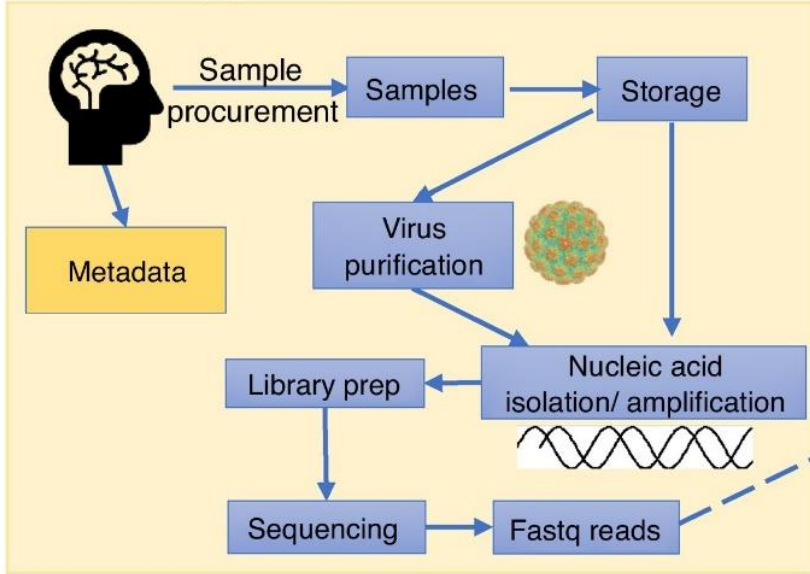
# Viral metagenomics

- Huge diversity!
- We can't yet guess the number of virus species
- Viral genomes are frequently less conserved
- Identification needs other approach
  1. Subtract host reads first: except integration sites!
  2. Assembly of viral contigs, starting from viral gene database
  3. Tblastn to compare with viral database: less sensitive to variation

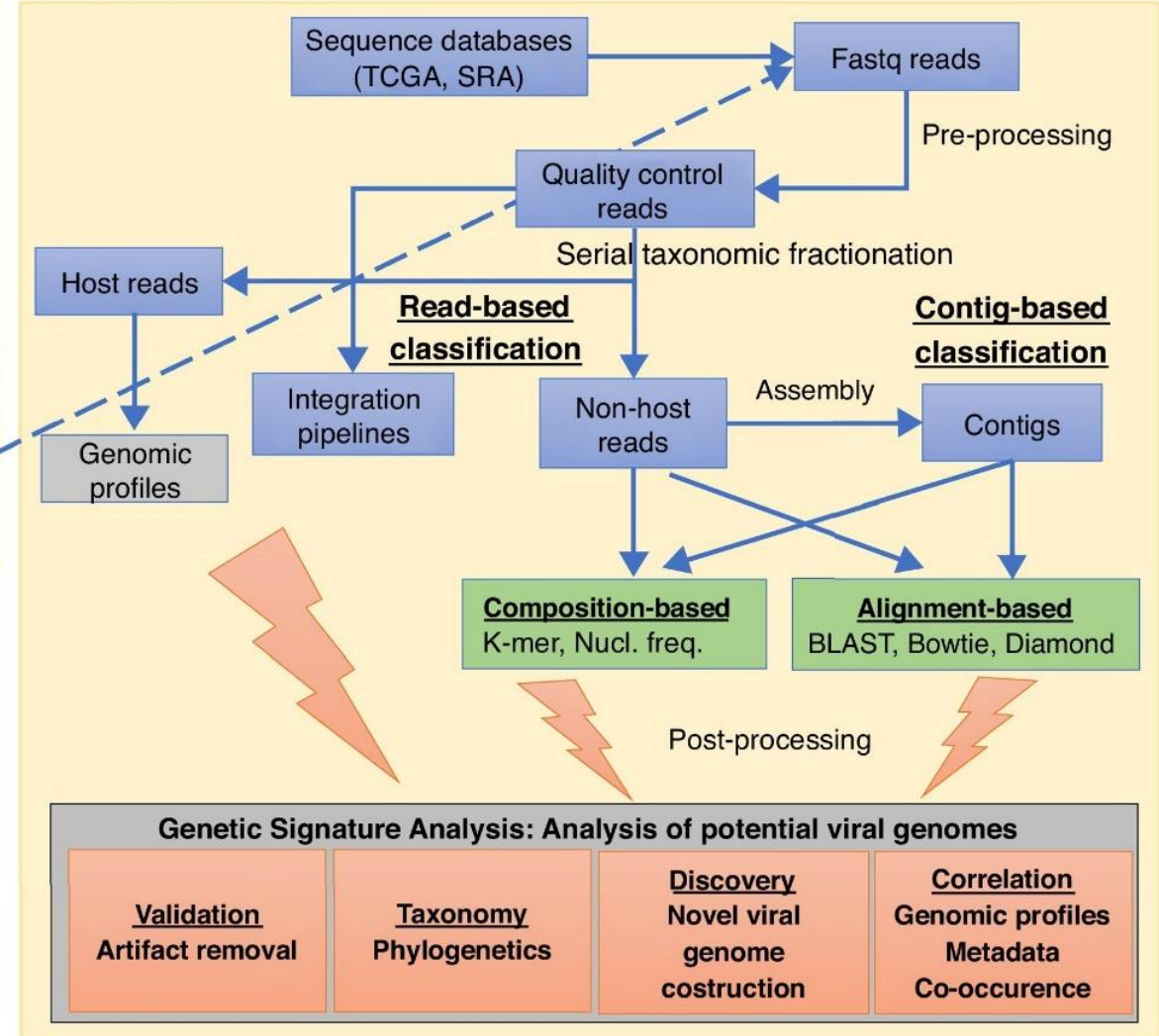
# Reducing search space in variable genomes



## Molecular pipeline

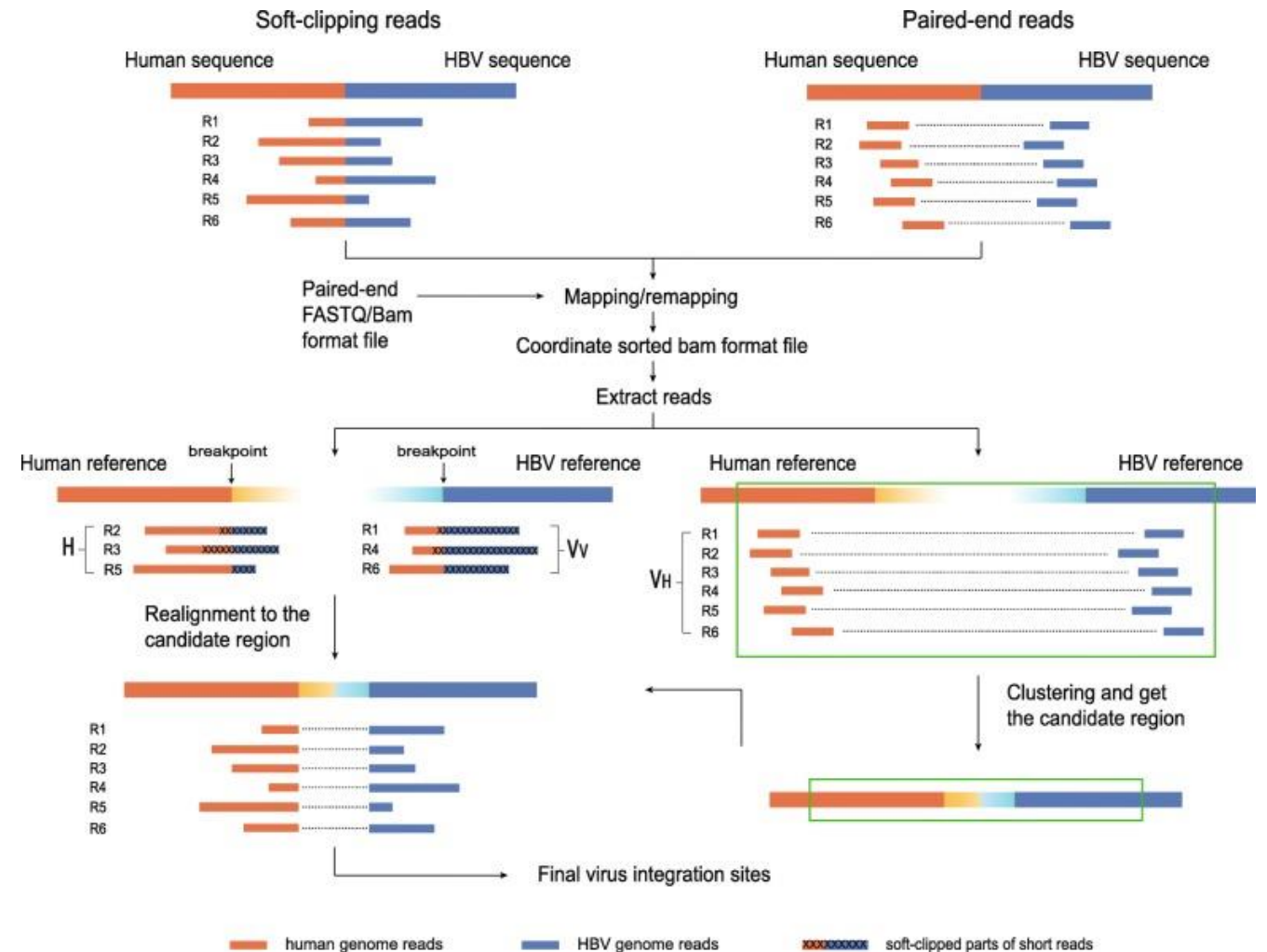


## Computational pipeline



# Viral integration site detection

- Sequence tissues with expected viral integration
- Use split-read info and repetitive softclip-mapping cycles
- Determine insertion site



# Contamination checks

- Purified/cultured samples are sometimes contaminated
- Metagenomics tools added to existing NGS pipelines:

What other DNA do we have in here?

- Quality control for entire workflow
- Early warning system for contamination
- Potential reduction in contamination cultures

# Making it difficult

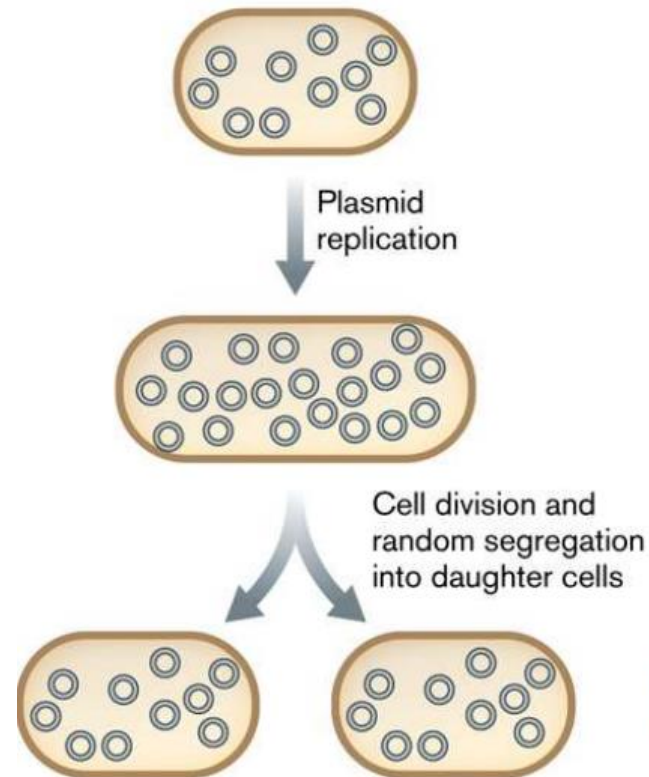
- Repetitive elements
  - Difficult to sequence
  - Low mappability
  - Problematic for *de novo* assembly



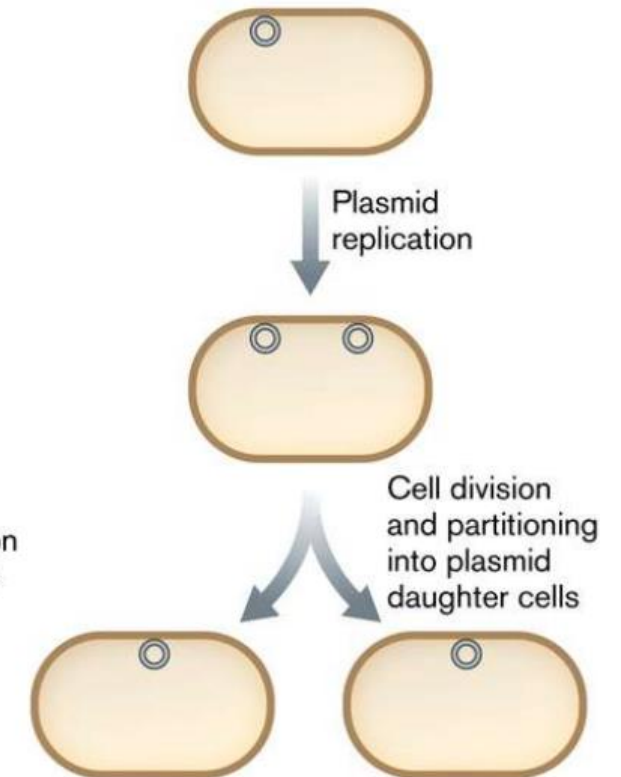
# Making it difficult

- Repetitive elements
- Plasmids
  - Abundance variable
  - Presence variable

A. For high-copy-number plasmids, random partitioning occurs.

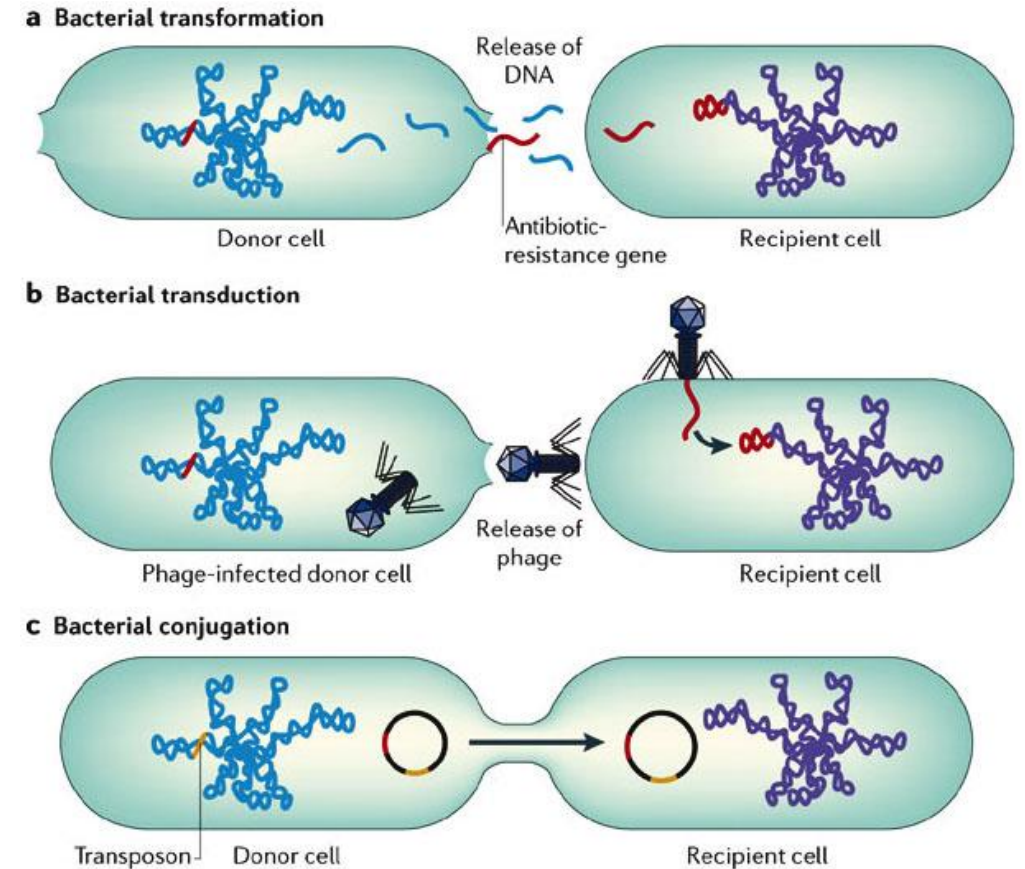


B. For low-copy-number plasmids, replication is coordinated with chromosome replication.



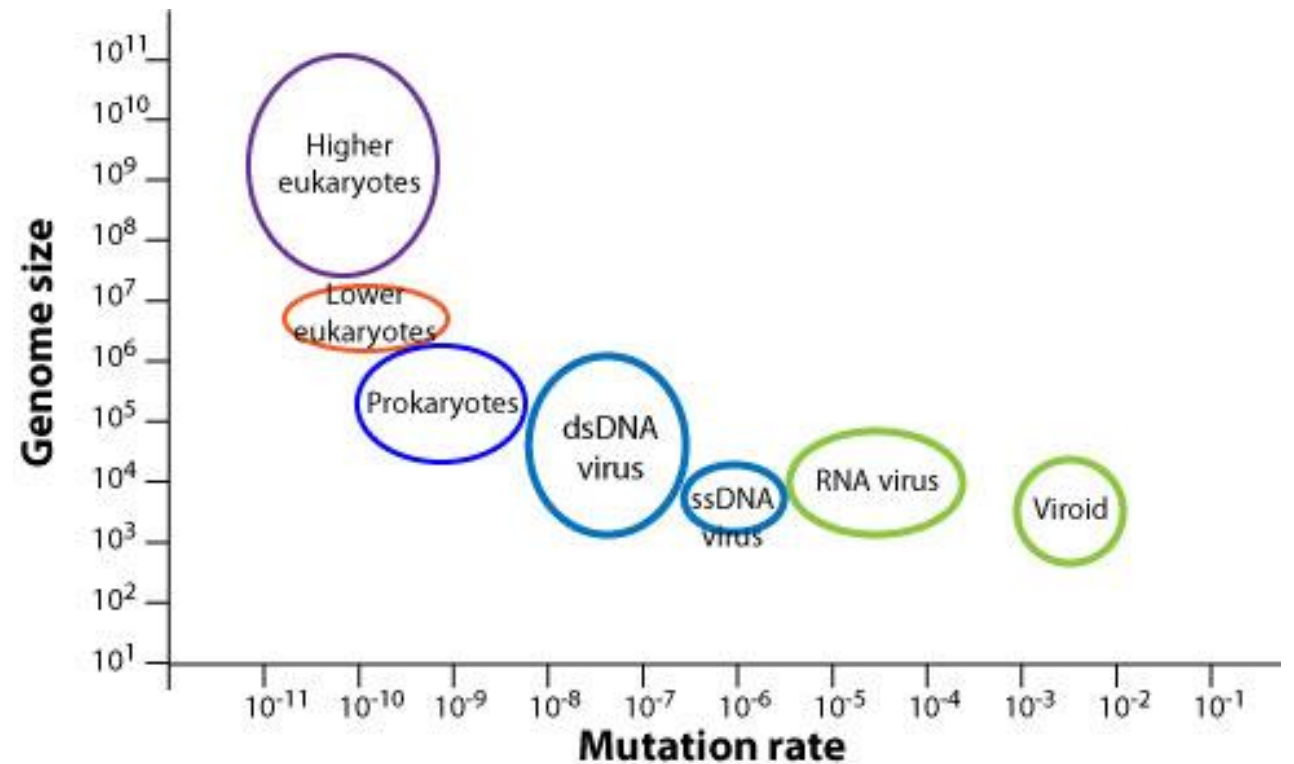
# Making it difficult

- Repetitive elements
- Plasmids
- Horizontal gene transfer
  - Transformation/conjugation/transduction
  - Species identification more difficult
  - Phylogeny gives conflicting results
  - Route of transmission difficult to trace

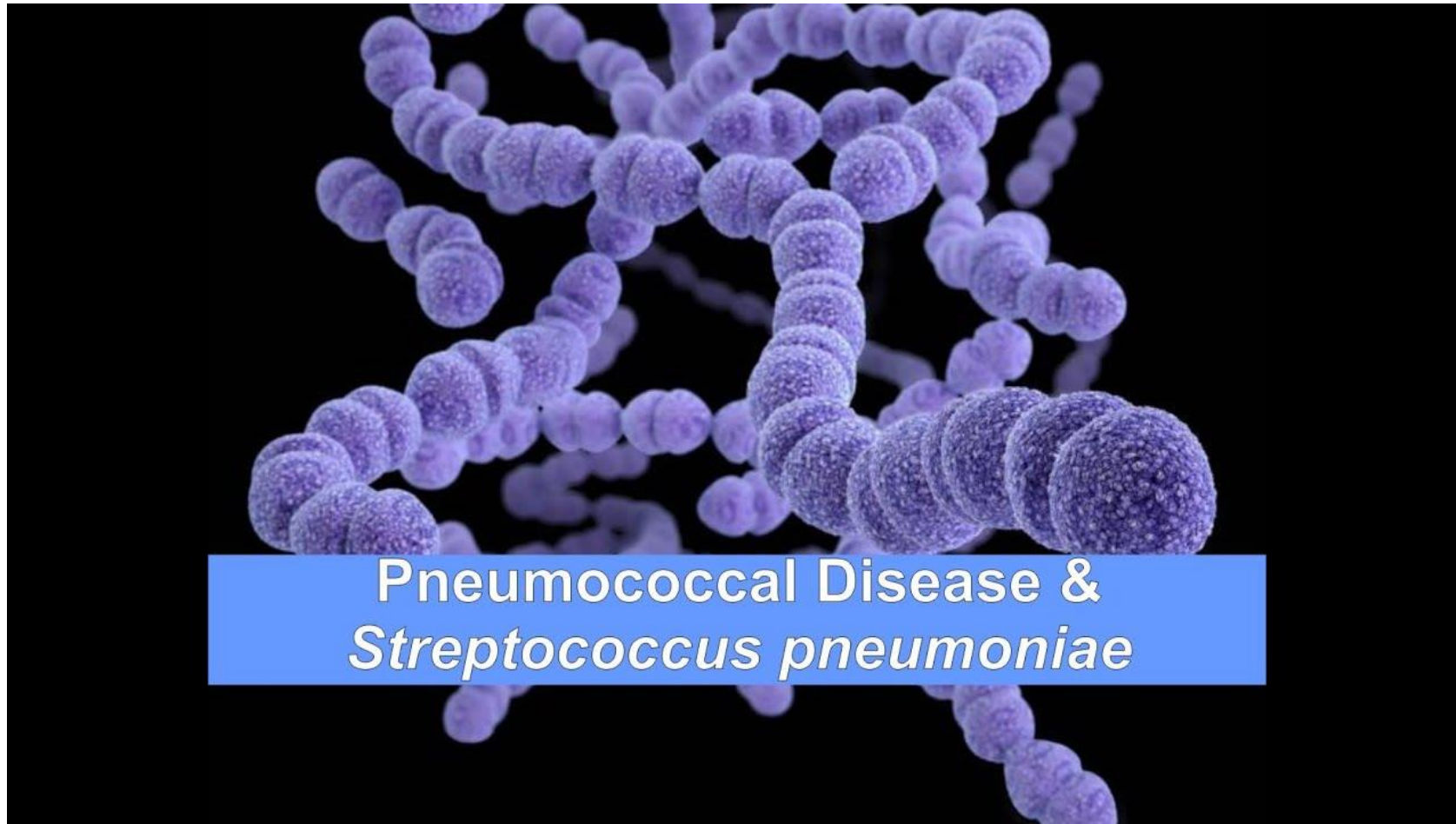


# Making it difficult

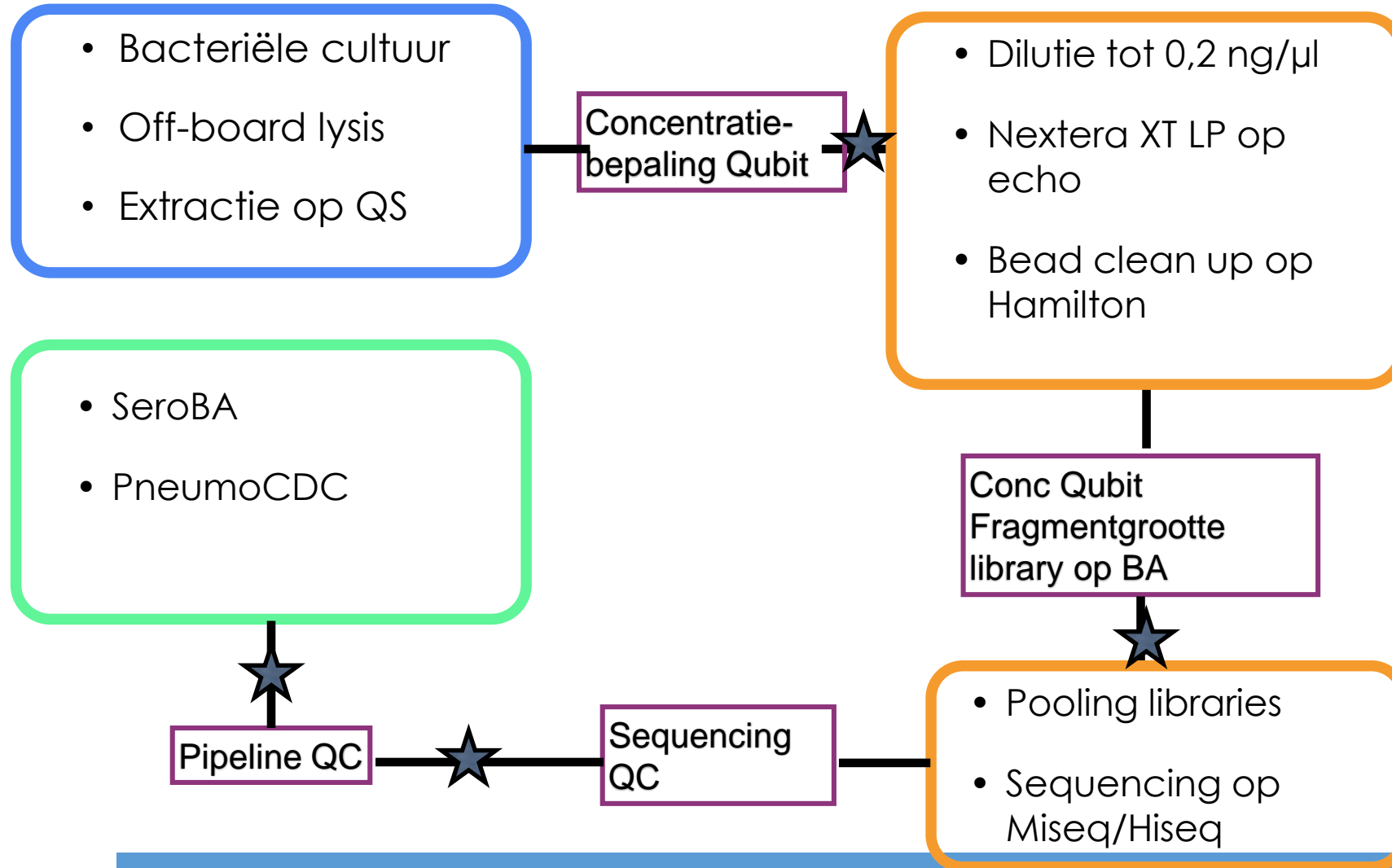
- Repetitive elements
- Plasmids
- Horizontal gene transfer
- High recombination/mutation rates



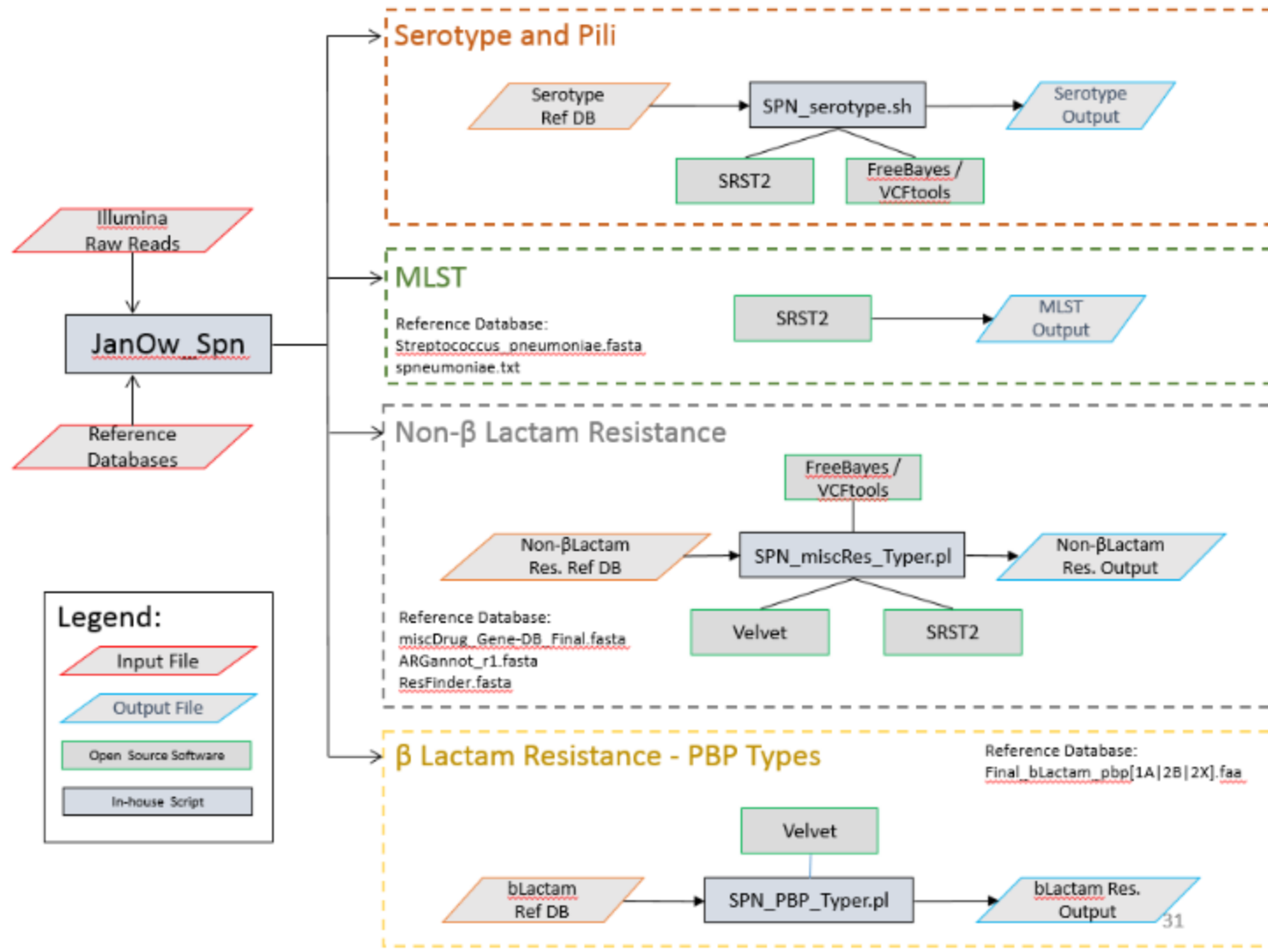
# Practical example



# SPN Workflow



# CDC pipeline - Sero/Pili, MLST, AB resistance



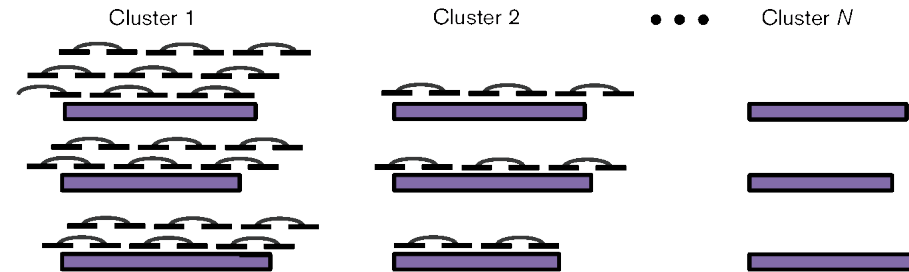
Metcalfe BJ et al, Clin Microb Infect. 2016; 22(1)

# SeroBA - Serotyping

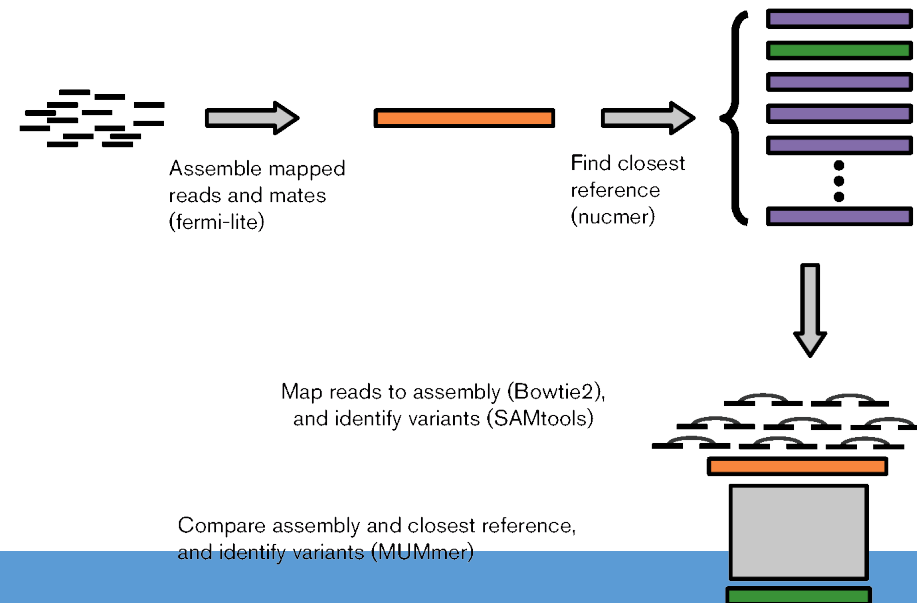
ARIBA  
Antimicrobial Resistance  
Identification By Assembly

mapping/alignment and targeted  
local assembly approach

Cluster reference sequences (cd-hit-est)  
and map all read pairs (minimap):



For each cluster that has reads mapped:

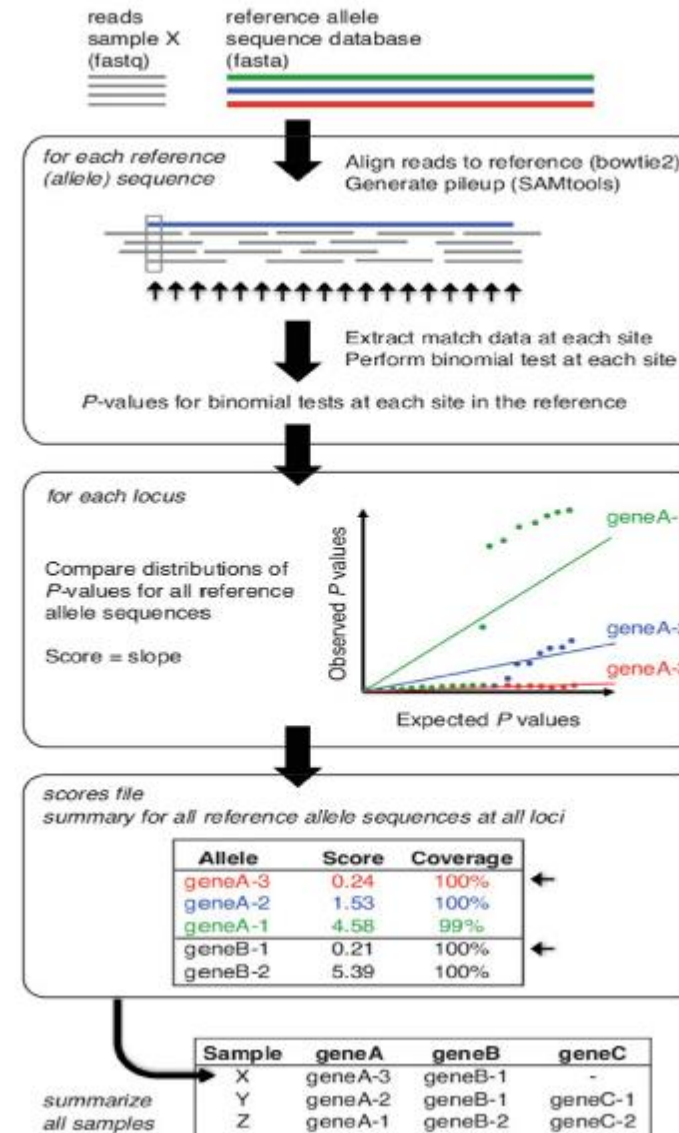
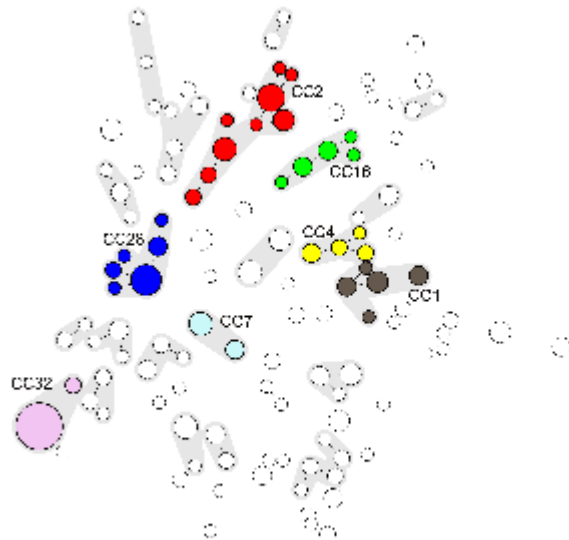


# MLST typing

Isolate characterization by multilocus sequence typing

Based on 7 housekeeping genes

SRST2 Short Read Sequence Typing for Bacterial Pathogens





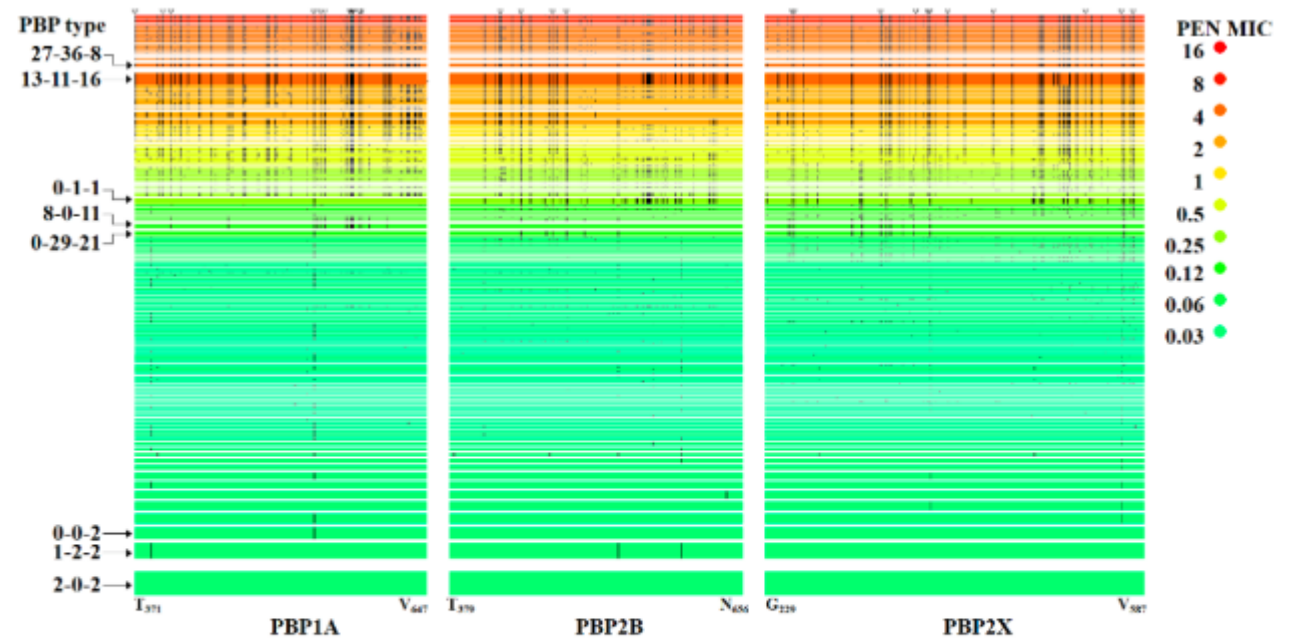
# Antimicrobial resistance – $\beta$ -Lactam AB

Mosaic genes – de novo assembly

Random forest model - PBP genes to MIC

	S	I	R
WGS_PEN_SIR_Meningitis	$\leq 0.06$	-	$\geq 0.12$
WGS_PEN_SIR_Nonmeningitis	$\leq 2$	4	$\geq 8$
WGS_AMO_SIR	$\leq 2$	4	$\geq 8$
WGS_MER_SIR	$\leq 0.25$	0.5	$\geq 1$
WGS_TAX_SIR_Meningitis	$\leq 0.5$	1	$\geq 2$
WGS_TAX_SIR_Nonmeningitis	$\leq 1$	2	$\geq 4$
WGS_CFT_SIR_Meningitis	$\leq 0.5$	1	$\geq 2$
WGS_CFT_SIR_Nonmeningitis	$\leq 1$	2	$\geq 4$
WGS_CFX_SIR	$\leq 0.5$	1	$\geq 2$

PEN: penicillin; AMO: amoxicillin; MER: meropenem  
 TAX: cefotaxime; CFT: ceftriaxone; CFX: cefuroxime



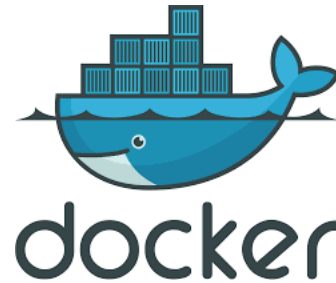
Li, Y. *et al.*, MBio. 2016 Jun 14;7(3). pii: e00756-16.

Li, Y. *et al.*, BMC Genomics. 2017 Aug 15;18(1):621.

Metcalf, BJ. *et al.* Clin Microbiol Infect. 2016 Jan;22(1):60.e9-60.e29.

# Automated cloud computing

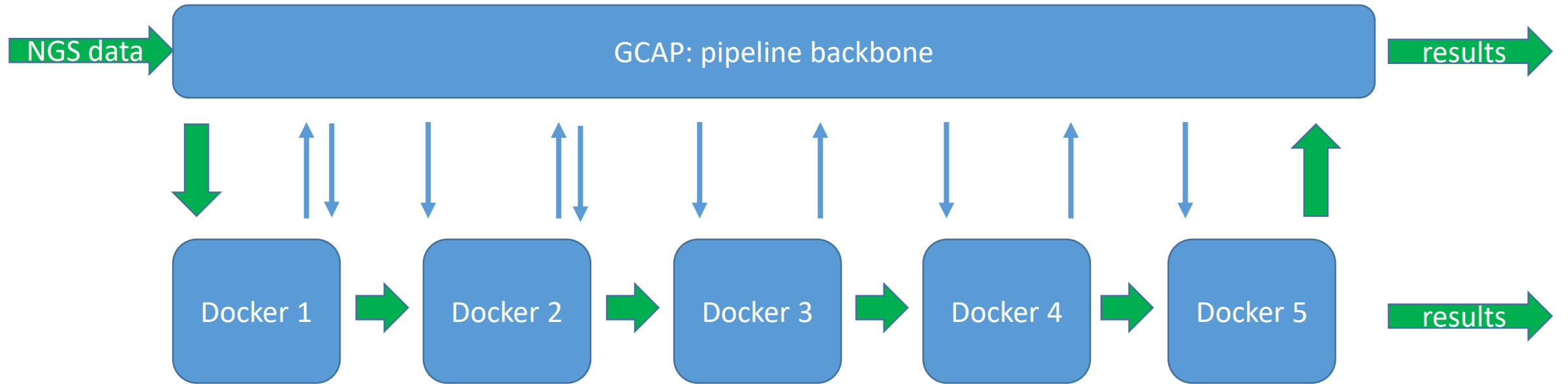
- Bring pipeline to the cloud using Docker



Docker is an open platform for developing, shipping, and running applications.

- Automated transfer of data upon sequencing
- Initiates different docker modules in the cloud
- Data and results stored and shareable through the cloud

# Cloud computing approach



# Case: *Streptococcus pneumoniae*

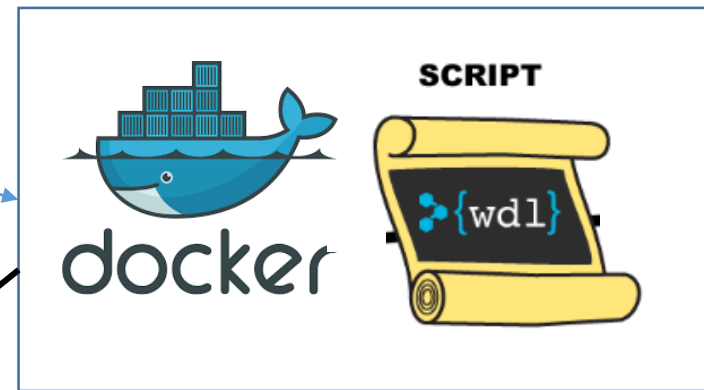
NGS goal: diagnosis, typing, susceptibility testing

Paper+  
GitHub distribution



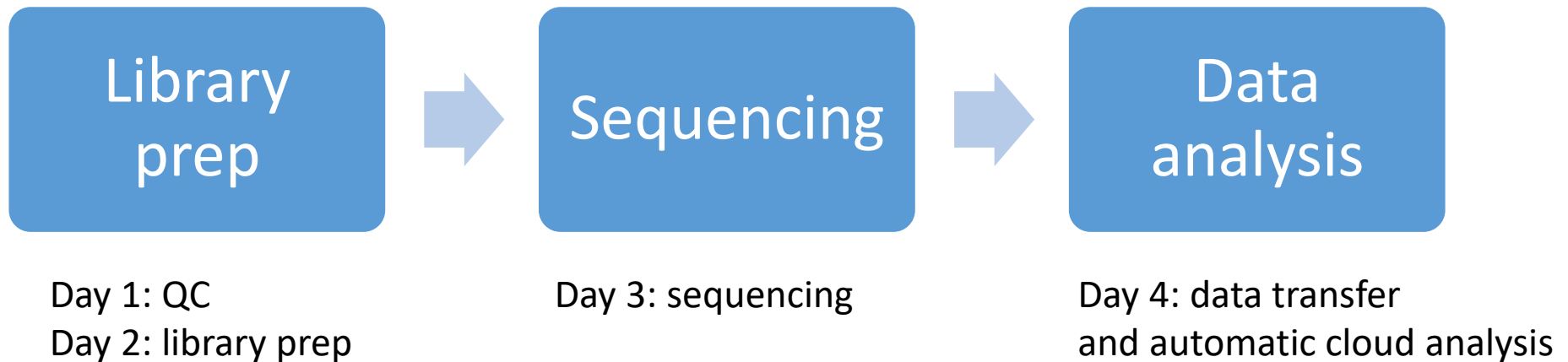
Labs worldwide

GENOMICS  
CORELEUVEN



# Timeline and cost reduction

- Library prep cost: 80% cost reduction by Echo use for library prep
- Sample number is scalable



# Benefits of NGS in microbiology

- Less need to grow bacteria
- Reduction in diagnostic time possible
- Get more and wider information
  - More detailed information
    - Species identification
    - Typing
    - Resistance gene
  - Ability to trace route of infection